

Cat.#R1100 **CleanWestern** Detection Kit  
(Membrane blocking step is not required)

Kit Components:

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|--|------------|
| 1. <b>Digital</b> Anti-mouse IgG-HRP (1mL)               | cat.#R1005 |
| 2. <b>Digital</b> Anti-Rabbit IgG-HRP (1mL)              | cat.#R1006 |
| 3. <b>KwikQuant</b> Ultra HRP Substrate Solution (100mL) | cat.#R1002 |
| 4. <b>CleanWestern</b> Buffer (3.5L)                     | cat.#R2001 |

Kit storage conditions and expiration:

Stability at 4°C: 24 months

## Procedures

1. Submerge the membrane in **CleanWestern** Buffer, and add your primary antibody directly in the buffer. Incubate with shaking 1-12 hours at room temperature. (Note: Blocking is not necessary when **CleanWestern** Buffer is used.)
2. Rinse the membrane with dI water twice.
3. Wash the membrane using **CleanWestern** Buffer 30 minutes once.
4. Dilute (@1:1000) the **Digital** HRP-secondary antibody in **CleanWestern** Buffer. Probe the membrane with rocking at room temperature for 1 hour.
5. Rinse the membrane with dI water twice.
6. Wash the membrane using **CleanWestern** Buffer for 30 minutes once.
7. Rinse the blot with de-ionized water\* briefly to remove the residual **CleanWestern** Buffer, and transfer the blot onto the sample plate of **KwikQuant** Imager. (\*The buffer contains reagents that will inhibit the reaction of HRP-ECL substrate.)
8. Submerge the membrane in 1:5 H<sub>2</sub>O-diluted **KwikQuant** Digital-ECL mixture (A:B=1:1) for 2 minutes.
9. Put the membrane on a **KwikQuant** Imager (or any chemiluminescence imagers) for image acquisition.
10. If the bands are too weak at 2 min exposure, submerge the membrane in non-diluted **KwikQuant**

Digital-ECL mixture (A:B=1:1) for another 2 minutes, followed by image acquisition.